AMENDMENTS TO THE SPECIFICATION

Please delete the paragraph on page 10, line 31, to page 11, line 14, and replace it with the following amended paragraph:

FIGURE 7. Sequence fDEC-205. (A) Schematic representation of DEC-205. (B) The predicted amino acid sequence of DEC-205 (SEQ ID NO:3) is aligned with the sequences of the bovine PLA2 receptor (SEQ ID NO:4) and the human macrophage mannose receptor (SEQ ID NO:5). Amino acid positions where there is identity among all three proteins are shaded. Protein domains are separated, and consensus amino acids that define C-type CRDs (Weis et al., Science 254:1608-15) are indicated blow the relevant sequence as follows: invariant amino acids are shown in single letter code, θ = aliphatic, χ = aliphatic or aromatic, Φ = aromatic, Z= E or Q, B = D or N, Ω = D, N, E, or Q. The two missing cysteines in CRD 8 are highlighted with a *. **The consensus sequences 'ΦθGθΩΩ', 'EΩCθ ', 'ΦθGθ', 'ZPBB', 'ΦθGθΩ' and 'EΩCθχ' are disclosed as SEQ ID NOS 7-12, respectively.** Peptide sequences determined by automated Edman degradation from purified DEC-205 protein are overlined and numbered (N indicates amino terminal, T indicated peptides generated with Trypsin, and L indicates peptides generated with endoproteinase lys-C). (C) Comparison of carboxyl-terminal cytoplasmic domain sequences of human (*top*) (SEQ ID NO:1) and murine (*bottom*) (SEQ ID NO:6) DEC-205. Regions of identity are underlined; regions of similarity are italicized.

Please delete the paragraph on page 56, lines 12-23, and replace it with the following amended paragraph:

Polyclonal antibodies to the N-terminal peptide-- The hapten-coupling strategy focused on the lone cysteine at residue 19 (Figure 6A). Peptide N1 (SESSGNDPFTIVHENTGKC) (SEQ ID NO: [[2]]**13**) was coupled to keyhole limpet hemocyanin (KLH) and ovalbumin (OVA) using maleimide chemistry (Imject, Pierce). An average of about 250 peptides were conjugated to each molecule of KLH, and about 6 peptides per molecule of OVA. The KLH-peptide conjugate was divided into aliquots of 400-500 μg each, and was injected eight times into two New Zealand White rabbits (200-250 μg per injection), again emulsifying into CFA for the initial immunization and IFA for boosts. To remove any anti-KLH reactivity from the sera, they were precleared on a KLH-cysteine column. Anti-peptide antibodies were isolated on a peptide-OVA column, where the peptide was coupled to an irrelevant carrier.